

# Method development and validation for simultaneous estimation of Fosnetupitant and Palonosetron by RP-HPLC

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## ABSTRACT

A simple and precise method was developed for estimating fosnetupitant as well as palonosetron. The method was found to be specific and precise. The separation was attained on Xterra RP18 column and linearity was achieved in the concentration rage of  $117\mu$ g/mL to  $470\mu$ g/mL of Fosnetupitant,  $0.125\mu$ g/mL to  $0.5\mu$ g/mL of Palonosetron with correlation coefficient 0.99. The percent recovery from the assay was found to be 100.19% for fosnetupitant and 100.30% for palonosetron. Limit of detection and quantitation for fosnetupitant and palonosetron were within the acceptable range. From the stability studies, the percentage variation was less than 10.0% which is the desired criteria. Therefore, this method can be adopted to estimate fosnetupitant as well as palonosetron in other pharmaceutical formulations.

Keywords: fosnetupitant, palonosetron, HPLC, Method development, Validation.

#### INRODUCTION

Fosnetupitant is the pro-drug form of netupitant. Generally, 25% to 30% of patients with a diagnosis of cancer receive chemotherapy as a treatment modality and 70% to 80% of these patients undergoing chemotherapy treatment may experience nausea and vomiting as major side effects. Considered one of the most distressing side effects of chemotherapy, nausea and vomiting has an immense impact on the quality of life of patients receiving certain antineoplastic therapies. Fosnetupitant is taken as an alternative treatment option for patients experiencing chemotherapy-induced nausea and vomiting.<sup>1</sup> Fosnetupitant chloride hydrochloride is white to off-white to yellowish solid or powder. Its solubility is pH dependent: at acidic pH (pH 2), its solubility is 1.4 mg/mL; at basic pH (pH 10), its solubility is 11.5 mg/mL.The fosnetupitant in this drug combination is a selective P/neurokinin-1 (NK-1) receptor antagonist<sup>2-4</sup>. Netupitant, the active moiety of fosnetupitant, is a selective neurokinin 1 (NK1) receptor antagonist with antiemetic activity. Netupitant competitively binds to and blocks the activity of the human substance P/NK1 receptors in the central nervous system (CNS), inhibiting NK1receptor binding of the endogenous tachykinin neuropeptide substance P (SP), which results in the prevention of chemotherapy-induced nausea and vomiting (CINV).

Palonosetron (INN, trade name Aloxi) is an antagonist of 5-HT3 receptors that is indicated for the prevention and treatment of chemotherapy-induced nausea and vomiting (CINV). It is the most effective of the 5-HT3 antagonists in controlling delayed CINV nausea and vomiting that appear more than 24 hours after the first dose of a course of chemotherapy and is the only drug of its class approved for this use by the U.S. Food and Drug Administration. As of 2008, it is the most recent 5-HT3 antagonist to enter clinical use.<sup>5-10</sup>Palonosetron hydrochloride is a white to off-white crystalline powder. It is freely soluble in water, soluble in propylene glycol, and slightly soluble in ethanol and 2propanol.Palonosetron is a 5-HT3 receptor antagonist with a strong binding affinity for this receptor and little or no affinity for other receptors. Cancer chemotherapy may be associated with a high incidence of nausea and vomiting, particularly when certain agents, such as cisplatin, are used. 5HT3 receptors are located on the nerve terminals of the vagus in the periphery and centrally in the chemoreceptor trigger zone of the area postrema.<sup>11-12</sup> Chemotherapeutic agents produce nausea and vomiting by stimulating the release of serotonin from the enterochromaffin cells of the small intestine. Serotonin then activates 5-HT3 receptors located on vagal afferents to initiate the vomiting reflex. The development of acute emesis is known to depend on

serotonin and its 5-HT3 receptors have been demonstrated to selectively stimulate the emetic response.

From the literature survey, it was revealed that few UV spectrophotometric method was developed but were not economical. Moreover, RP-HPLC<sup>13</sup> and LC-MS<sup>14</sup>and

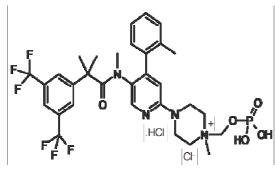


Figure 1: Structure of fosnetupitant

## **MATERIALS AND METHODS**

**Materials:** Gift samples of fosnetupitant and palonosetron were received from Startech lab, Hyderabad. KH2PO4 was purchased from Final chemicals whereas water, methanol for HPLC and ortho phosphoric acid were purchased from Merck.

**Instrumentation:** Waters HPLC was used for the separation of fosnetupitant and palonosetron. UV/VIS spectrophotometer (LABINDIA UV 12.500<sup>+</sup>) was used for detection. Instruments such as; pH meter used was of Adwa-AD 10100 and weighing machine was of Afcoset ER-1000A.

#### **Method Development**

**Preparation of Phosphate buffer:** Prepare 800 mL of distilled water in a suitable container. Add 20.214 g of  $Na_2HPO_4$ -7H<sub>2</sub>O to the solution. Add 3.394 g of  $NaH_2PO_4H_2O$  to the solution. Adjust solution to final desired pH of 7.4 using NaOH. It was then subjected to filtration through membrane filter followed by sonication for 10mins.

**Mobile phase preparation:** From the above prepared buffer, 700mL is mixed with 300mL of HPLC methanol, mixed, degassed, sonicated for 10min followed by filtration via vacuum filter.

**StockSolution Preparation:** The mixed stock solution was prepared freshly by dissolving 235 mg of fosnetupitant and 0.25 mg of palonosetron in 100 mL of mobile phase.

**Standard solution preparation:** 235 mg of fosnetupitant and 0.25 mg of palonosetronworking standard were weighed

derivative methods were also developed which estimates fosnetupitant and palonosetron either individually or in combination. In the present research work, a new method was developed to estimate fosnetupitant and palonosetron simultaneously and validated as per ICH guidelines.<sup>15</sup>

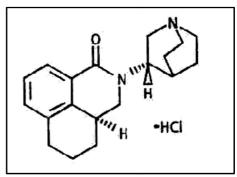


Figure 2: Structure of palonosetron

and added in volumetric flask of 100mL quantity. To it, small quantity of diluent was occasionally added and sonicated and made a final dilution to the mark of the VF using diluent and used as stock solution. From this, 1mL transferred to 10mL VF and diluted using diluents to obtain final volume of 10.0mL.

**Sample solution preparation:** Pipette 1mL of Fosnetupitant and Palonosetron of the above stock solution into a 10mL volumetric flask and dilute up to the mark with Diluents. To it, small quantity of diluent was occasionally added and sonicated and made a final dilution to the mark of the VF using diluent and used as stock solution.

**Procedure:** Mixture of buffer (pH 7.4) and methanol in the ratio 70:30% v/v was used as mobile phase which was injected into the system for 30 minutes prior to injecting the prepared solutions of standard as well as sample. Detection of the drug was achieved at the wavelength of 245nm at 25°C. After several trials, method was optimized followed by validation of the method considering various validation parameters.

## **RESULTS AND DISCUSSION**

Method development was achieved using Xterra RP18 (4.6 x 150mm, 5.0 $\mu$ m). Mobile phase was mixture of Phosphate buffer and methanol (70:30% v/v). Flow rate (1mL/min) and injection volume (20 $\mu$ l) was set. The peaks obtained had good resolution with the retention time 2.236 and 3.682 for fosnetupitant and palonosetron respectively. Chromatogram of optimized trial is shown in figure 3.

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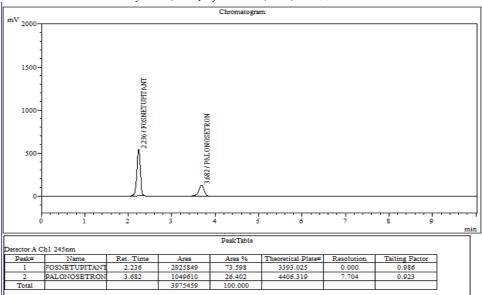


Figure 3: Chromatogram of optimized trial

**System suitability:** All the parameters were evaluated by performing system suitability studies. The recorded responses for suitability studies are depicted in table 1.

Table 1: Results of system suitability parameters

S.No	Name	RT (min)	Area (µV sec)	USP resolution	USP tailing	USP plate count
1	Fosnetupitant	2.236	2925849		0.986	3393.025
2	Palonosetron	3.682	1049610	7.704	0.923	4406.319

**Method validation:** Validation of the method was evaluated for various parameters which include linearity, specificity, robustness and stability. The method was also evaluated for specificity of the method and was found to be specific as there were no interactions found. Linearity obtained was shown to have good correlation as shown in table 2.

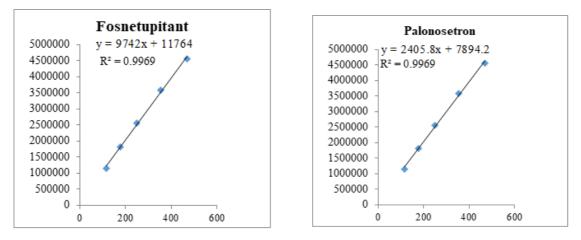
	Label Claim (mg)	% Assay
Fosnetupitant	235	100.19
Palonosetron	0.25	100.30

**Linearity:** The linearity range was observed from  $117\mu$ g/mL to  $470\mu$ g/mL of Fosnetupitant,  $0.125\mu$ g/mL to  $0.5\mu$ g/mL of Palonosetron. The respective absorbance

values are depicted in table 3. The linearity graph plotted is presented in figure 4 and 5 for Fosnetupitantas well as Palonosetron respectively.

#### **Table 3:Linearity results**

S. No.	Fosnetupitant		Palonosetron	
	Concentration (µg/mL)	Area	Concentration (µg/mL)	Area
1	117	1144247	0.125	386439
2	176	1812362	0.187	622922
3	235	3062738	0.250	1090765
4	352	4069834	0.375	1351741
5	470	4544205	0.500	1672640
	Slope (m)	9742	2405	
	Intercept (c)	11764	7894	
Corr	relation coefficient (R <sup>2</sup> )	0.9969	0.9972	



## Figure 4: Linearity graph for FosnetupitantFigure 5: Linearity graph for Palonosetron

Accuracy: Percent recovery of sample solutions at different concentrations (50%, 100%, and 150%) was calculated. The Percent recovery of fosnetupitant and palonosetron are depicted in table 4 and 5 respectively.

% Concentration	Area	Added amount (mg)	Amount Found (mg)	PercentRecovery	Mean Recovery
50%	1144247	117.0	117.22	100.18	
100%	3062738	235.0	235.46	100.19	100.17
150%	4544205	470	470.66	100.14	-

#### Table 4: Accuracy (recovery) data for Fosnetupitant

%Concentration	Area	Added amount (mg)	Amount Found (mg)	% Recovery	Mean Recovery
50%	386439	0.125	0.126	100.8	
100%	1090765	0.250	0.2512	100.3	100.5
150%	1672640	0.500	0.502	100.4	-

#### Table 5: Accuracy (recovery) data for Palonosetron

**Precision:** Precision of the method was performed for both sample solutions as described under experimental work. The same method was performed on the other day for intermediate precision. The results are depicted in the table 6 and 7.

#### Table 6: Results of precision

Injection	Area (Fosnetupitant)	Area (Palonosetron)
Injection-1	2855391	1024050
Injection-2	2853669	1023557
Injection-3	2930368	1049612
Injection-4	2916908	1041245
Injection-5	2925849	1049610
Average	2896437	1037615
<b>Standard Deviation</b>	20566	7723
%RSD	0.71	0.74

#### Table 7: Results of intermediate precision

Injection	Area (Fosnetupitant)	Area (Palonosetron)
Injection-1	2853414	1023822
Injection-2	2951094	1059836
Injection-3	2925582	1056103
Injection-4	2916466	1041407
Injection-5	2967660	964840

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Injection-6	2973564	1035268
Average	2931297	1030213
Standard Deviation	14319	5679
%RSD	0.48	0.55125

**Limit of Detection and Quantitation:** Lowest concentrations of the sample were prepared and measured the signal/ noise ratio. Chromatogram for LOD and LOQ is presented in figure 6 and 7 and the results are depicted in table 8.

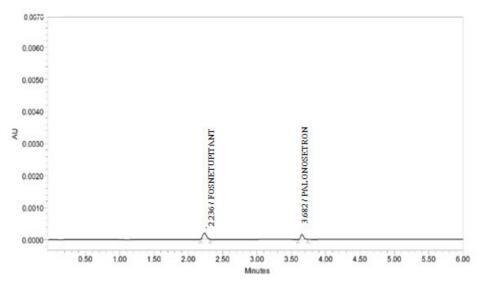


Figure 6: Chromatogram of Fosnetupitant, Palonosetron showing LOD

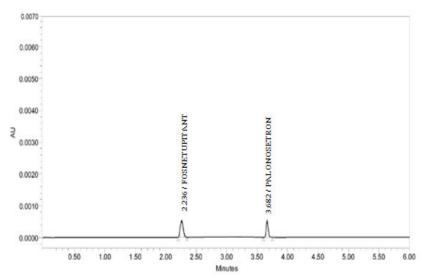


Figure 7: Chromatogram of Fosnetupitant, Palonosetron showing LOQ

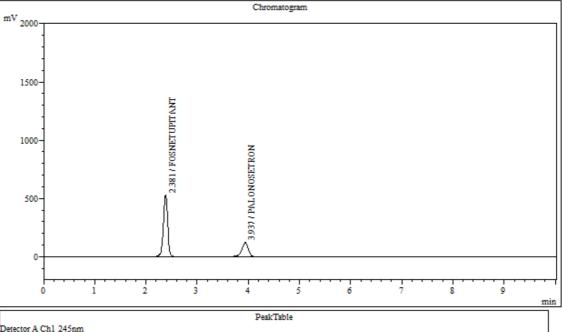
Drug name	Baseline noise (µV)	Signal obtained (µV)	S/N ratio
	Limit of d	etection	
Fosnetupitant	58	172	2.97
Palonosetron	58	170	2.93
	Limit of qu	antitation	
Fosnetupitant	58	578	9.97
Palonosetron	58	577	9.95

Table 8: Results of LOD and LOQ

**Robustness:** The standard and samples of both drugs were injected by changing the conditions of chromatography. There was no change observed in the parameters like tailing

factor, resolution, plate count and asymmetric factor. Chromatograms for variation in flow rate are presented in figure 8 and 9 whereas chromatograms for variation in

## Variation in flow



:h1 245nm			
Name	Ret. Time	Area	Area %
FOSNETUPITANT	2.381	3030699	74.127
PALONOSETRON	3.937	1057808	25.873
		4088507	100.000
	Name FOSNETUPITANT	Name Ret. Time FOSNETUPITANT 2.381	Name         Ret. Time         Area           FOSNETUPITANT         2.381         3030699           PALONOSETRON         3.937         1057808

### Figure 8: Chromatogram showing less flow

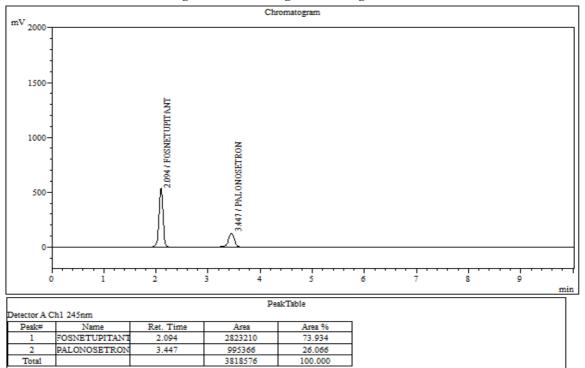
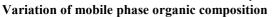
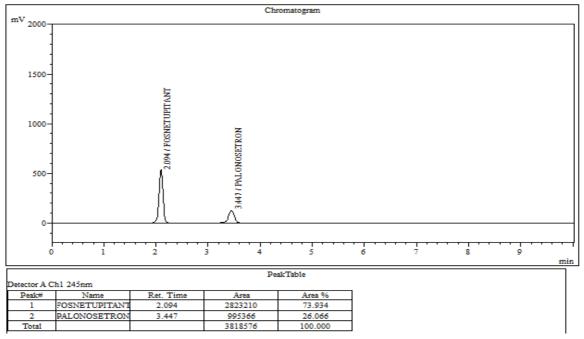
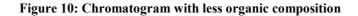


Figure 9: Chromatogram showing more flow

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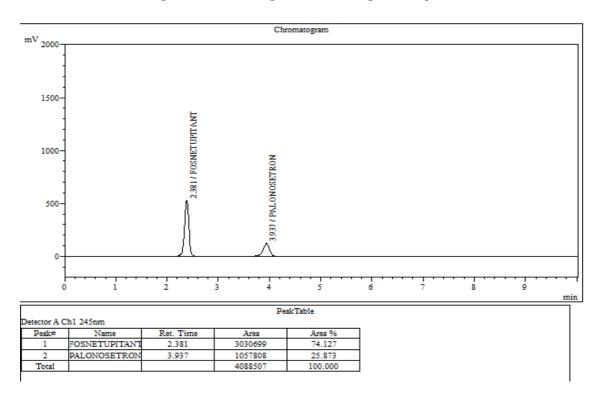


Figure 11: Chromatogram with more organic composition

Table 9: Results for variation in flow for Fosnetupitant
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S. No	Flow Rate (mL/min)	System Suitability Results	
		<b>USP Plate Count</b>	USP Tailing
1	0.9	2497.32	1.39
2	1.0	2491.44	1.39
3	1.1	2501.26	1.39

#### Arshiya *et al / Int. J. of Farmacia*, 2020; Vol-(6)3: 82-90 Table 10: Results for variation in flow for Palonosetron

S. No	Flow Rate (mL/min)	System Suitability Results	
		<b>USP Plate Count</b>	USP Tailing
1	0.9	2768.69	1.61
2	1.0	2736.33	1.61
3	1.1	2789.96	1.60

\* Actual flow (1.0mL/min) was considered from Assay standard.

Table 11: Results for variation in mobile phase composition for Fosnetupitant

S. No	Change in Organic Composition in the Mobile Phase	System Suitability Results	
		<b>USP Plate Count</b>	USP Tailing
1	10% less	2468.33	1.40
2	*Actual	2491.44	1.39
3	10% more	2465.45	1.41

S. No	Change in Organic Composition in the Mobile Phase	System Suitability Results	
		<b>USP Plate Count</b>	USP Tailing
1	10% less	2759.45	1.62
2	*Actual	2736.33	1.61
3	10% more	2726.34	1.61

\* Results for actual Mobile phase composition have been considered from Accuracy standard.

## CONCLUSION

From the above experimental results and parameters it was concluded that, this newly developed method for the simultaneous estimation ofFosnetupitant and Palonosetron was found to be simple, precise, accurate and high resolution and shorter retention time makes this method more acceptable and cost effective and it can be effectively applied for routine analysis in research institutions, quality control department in industries, approved testing laboratories, bio-pharmaceutical and bio-equivalence studies and in clinical pharmacokinetic studies in near future.

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